

Maintenance of *Arabidopsis thaliana* T87 cell suspension culture

Introduction

T87 cell suspension culture was obtained from a seedling of *Arabidopsis thaliana* L. (Heynh.) ecotype Columbia (Axelos *et al.* 1992). The culture is composed of small, near-uniform clumps of cells, has a green color (Figure1), and is maintained in Jouanneau and Péaud-Lenoël (JPL) medium under continuous light at 22°C and subcultured at two-week intervals.

Materials

I. Stock Solutions and Chemicals

A) JPL A' (1 L)	
KNO ₃	65.5 g
CaCl ₂ ·2H ₂ O	4.4 g
MgSO ₄ ·7H ₂ O	3.7 g
KH ₂ PO ₄	1.7 g
B) JPL B (1 L)	
H ₃ BO ₃	6.2 g
MnSO ₄ ·4H ₂ O	22.3 g
ZnSO ₄ ·7H ₂ O	10.6 g
KI	0.83 g
Na ₂ MoO ₄ ·2H ₂ O	0.25 g
CoCl ₂ ·6H ₂ O	0.025 g
CuSO ₄ ·5H ₂ O	0.025 g
C) JPL C (1 L)	
FeSO ₄ ·7H ₂ O	2,780 mg
Na ₂ -EDTA	3,730 mg
D) JPL D (1 L)	
<i>myo</i> -Inositol	10 g
Glycine	0.2 g
E) JPL VT (100 mL)	
Nicotinic acid	50 mg
Pyridoxine·HCl	50 mg
Thiamine·HCl	40 mg
F) JPL P (100 mL)	
200 mM KH ₂ PO ₄	19.5 ml
200 mM Na ₂ HPO ₄	30.5 ml
H ₂ O	50 ml

- G) 1 mM NAA (100 mL)
1-Naphthaleneacetic acid 18.62 mg
- H) Casein hydrolysate, vitamin-free
Casamino acids vitamin assay, Difco (228820)

II. Glassware and Stainless Sieves

(All are sterilized by autoclaving at 121°C for 20 min.)

- A) Erlenmeyer flask (300 ml), capped with two layers of aluminum foil
- B) Pipette, large tip opening (10 ml), and a bulb
- C) Stainless sieve (diameter, 5 cm; pore size, 1 mm) set on a tall beaker (200 ml), capped with two layers of aluminum foil

III. Preparation of JPL Medium

- A) Prepare three solutions as follows:

A)-1 JPL mineral solution (1 L)

- JPL A' 37.5 ml
- JPL B 0.375 ml
- JPL C 2.5 ml
- Adjust pH to 5.7 with 0.2 N KOH.

A)-2 JPL organic solution (100 mL)

- Casein hydrolysate 0.1 g
- JPL D 10 ml
- JPL VT 1 ml
- Adjust pH to 5.7 with 0.2 N HCl.

A)-3 JPL sucrose solution (100 mL)

- Sucrose 15 g
- JPL P 1 ml
- 1 mM NAA 1 ml

- B) Autoclave these solutions at 121°C for 20 min.
- C) Add 800 ml of JPL mineral solution, 100 ml of JPL organic solution, and 100 ml of JPL sucrose solution aseptically.
- D) Pour 80 ml of JPL medium into a sterile flask.

Methods

- I. Filter a two-week-old cell suspension through a stainless sieve (Figure 2).
- II. Agitate the filtrate well and transfer 2 ml of cell suspension to 80 ml of fresh JPL medium with a pipette.
- III. Incubate cell cultures on a rotary shaker at 120 rpm under continuous light (40–100 $\mu\text{E/s/m}^2$) at 22°C.

Notes

- I. T87 cells may be maintained in other media. We have culture them in JPL medium as described in the original report by Axelos *et al.* (1992). Yamada *et al.* (2004) reported that T87 cells grew well in Gamborg's B5 medium. When T87 cells are cultured in the medium other than JPL medium, it is necessary to optimize the subculture method, *e.g.*, subculture interval, amount of cells transferred, and the pore size of the sieve.
- II. T87 cells should be transferred to fresh medium immediately after arrival. We transport T87 cells on semi-solid JPL medium in 250-ml disposable flasks. T87 cells should be collected from the medium with a spatula and transferred to Erlenmeyer flasks containing fresh liquid medium. The cell suspension culture should be carefully established from small scale fractions of the original culture (*e.g.*, 20 ml of medium in a 100-ml flask) because some T87 cells may have been damaged during transport.
- III. The density of living cells subcultured to fresh medium is important for stable maintenance of cell suspension cultures. A high initial density causes overgrowth after the usual culture period, while a low density suppresses cell division. Occasionally, T87-cell clumps develop into large aggregates, which cause a decrease in the number of cells passed through a 1-mm sieve. In such cases, it should be confirmed that an adequate amount of cells are transferred to each fresh medium during subculturing.
- IV. Under our culture conditions, the mass of T87 cells had increased around 50-fold by day 14 of culturing (Figure 3). In addition, T87 cells proliferated in the dark.

References

- Axelos M, Curic C, Mazzolini L, Bardet C, Lescure B (1992) A protocol for transient gene expression in *Arabidopsis thaliana* protoplasts isolated from cell suspension cultures. *Plant Physiology and Biochemistry* 30: 123-128
- Yamada H, Koizumi N, Nakamichi N, Kiba T, Yamashino T, Mizuno T (2004) Rapid response of *Arabidopsis* T87 cultured cells to cytokinin through His-to-Asp phosphorelay signal transduction. *Bioscience, Biotechnology, and Biochemistry* 68: 1966-1976

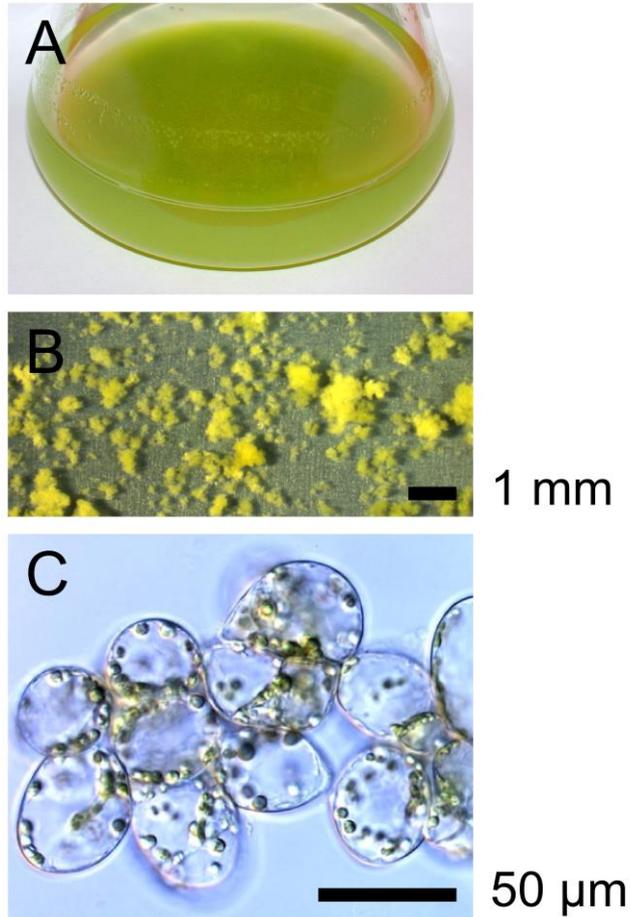


Figure 1 *Arabidopsis thaliana* T87 cell suspension culture

- (A) Two-week-old cell suspension culture.
- (B) Cell clumps. The size of cell clumps are varied, but most of them are below 1 mm.
- (C) Microscopic observation of T87 cells. The cells contain many chloroplasts.



2-week-old T87 suspension culture in a 300-ml flask.



Pass the cell suspension through a stainless sieve.

Stainless sieve (pore size, 1 mm)



Transfer 2 ml of filtrate to 80 ml of fresh JPL medium.

Figure 2 Procedure for subculturing T87 cells

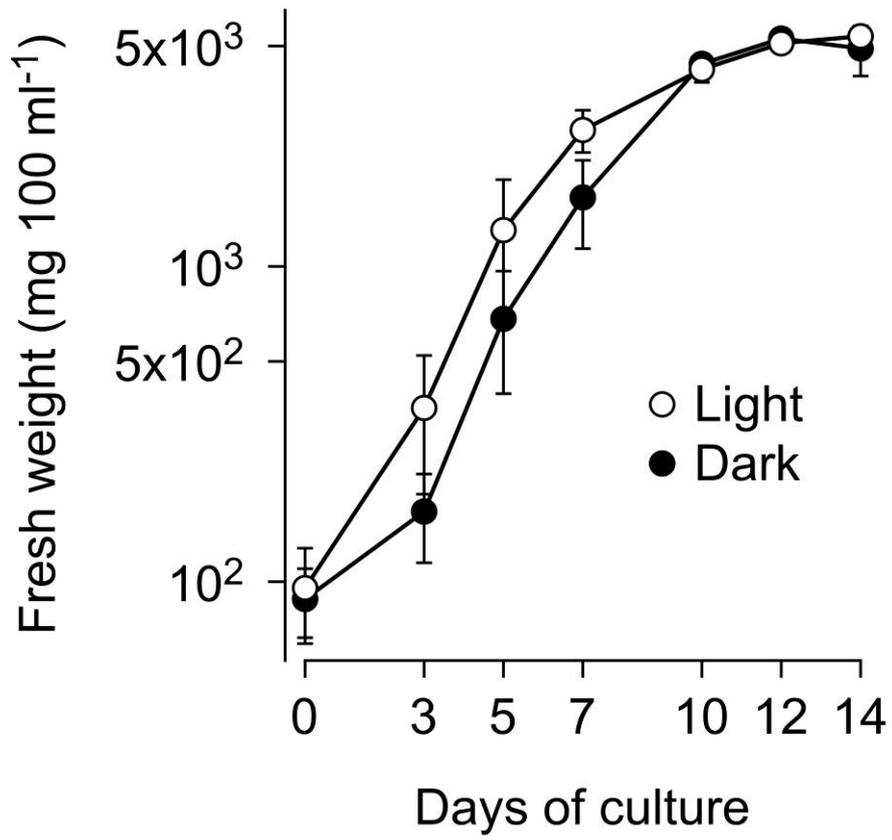


Figure 3 Growth profiles of T87 cells